CHEMICAL STRUCTURE OF THE ETHER LIPIDS OF THERMOPHILIC ACIDOPHILIC BACTERIA OF THE *CALDARIELLA* GROUP

Mario de Rosa,* Salvatore de Rosa,* Agata Gambacorta,* Luigi Minale* and John D. Bu'Lock†

*Laboratorio per la Chimica M. I. B. del C. N. R. Via Toiano 2, Arco Felice, Napoli, Italy; †Department of Chemistry, University of Manchester, England

Abstract—The lipids of the *Caldariella* group of extremely thermophilic acidophilic bacteria are based on a 72-membered macrocyclic tetraether made up from two C_{40} diol units and either two glycerol units or one glycerol and one nonitol. The C_{40} components have the 16,16'-biphytanyl skeleton and the detailed structure of three of them is established.

INTRODUCTION

The prevalence of lipids based on isoprenoid glyceryl ethers in very thermophilic acidophilic bacteria such as *Sulfolobus*, *Thermoplasma*, and others, classified by us as the *Caldariella* group, [1-3] has been established as a characteristic of the whole group [4] and our preliminary characterisation of the major components as a mixture of cyclic ethers each combining a 2,3-glyceryl unit etherified with a C_{40} isoprenoid residue has been briefly described [4,5]. Since cleavage of the mixture with BCl_3 gave glycerol and the C_{40} dichlorides, $C_{40}H_{76-80}Cl_2$, the cyclic ethers were formulated as (1). Preliminary evidence was also presented to show that the C_{40} residues were formally derived by ω,ω'-linkage (i.e. head-to-head) of two O-phytanyl residues, with or without further cyclisations, giving an acyclic, a monocyclic, and a bicyclic C_{40} component; this conclusion was based on NMR and mass spectroscopy data obtained for the C_{40} diols and the corresponding hydrocarbons.

In this paper we report evidence that one of the two major series of ether lipids (obtained after hydrolysis of the total complex lipids) is to be formulated not as (1) but as (2) i.e. as a 72-membered macrocyclic tetraether combining two glycerols and two C_{40} residues, a conclusion which has also been reached independently by Langworthy [6]. We also report detailed structural assignments for the three types of C_{40} residue contained in the lipids of this series. These diglycerol di(biphytanyl) tetraethers are accompanied by a second lipid class, in which one of the two glycerols is replaced by a nonitol and the biphytanyl residues are more extensively cyclized; detailed description of this second series will be given in a later account.

RESULTS

The macrocyclic tetraether structure

Determinations of the molecular weight by vapour pressure osmometry of the mixed diglycerol tetraethers in $CHCl_3$ gave a value of M = 1290 [(1) requires 646–650; (2) requires 1292–1300], and for corresponding diglycerol tetraether diacetate mixture M = 1350. In NMR spectra of the tetraether diacetate the signal due to the —CH_2OAc protons (δ 4.14, multiplet) is coupled to the alkoxy methine proton signal (δ 3.6, complex and overlapping with alkoxymethylene signal) and this established structure (2) for the diglycerol tetraethers (without however distinguishing between the alternative attachments of the second carbinol group). Similar but more preliminary data on the glycerol nonitol tetraethers indicate an analogous structure, with the nonitol replacing one of the two glycerols in (2).

The C_{40} ω,ω'-biphytanyl components

Information on the isoprenoid structures was obtained mainly by detailed GC–MS studies on the derived hydrocarbons and by ^1H NMR; using the europium shift reagent on the separate C_{40} diols and on C_{36} and C_{32} diols derived by chain-shortening reactions.

The C_{40} hydrocarbons

Conversions of the tetraethers into the C_{40} di-iodides with HI and of the diiodides into the C_{40} hydrocarbons by $LiAlH_4$, have already been described as has the GLC separation of the hydrocarbon mixture. Hydrocarbons with —$C^1H_2{}^2H$ terminal groups were also prepared (by using $LiAl^2H_4$) and these $^2H_2$-hydrocarbons were useful in assigning certain ions in the mass spectra. Whereas the diglycerol tetraethers afforded the hydrocarbons $C_{40}H_{78}$, $C_{40}H_{80}$, and $C_{40}H_{82}$, discussed in detail below, the corresponding hydrocarbons from the glycerol nonitol tetraethers covered the range $C_{40}H_{74}$, $C_{40}H_{76}$, and $C_{40}H_{78}$, the last being identical with the bicyclic hydrocarbon from the diglycerol tetraether mixture.

The MS of the saturated acyclic hydrocarbon $C_{40}H_{82}$ (M$^+$ m/e 562; $^2H_2$ analogue, 564) can be rationalized as in Scheme 2 in terms of (a) a series of cleavages, both with and without H transfer to the uncharged fragment, α to >CHMe groups and removing successively 7,5,5,4 and 5C atoms, finally generating the base peaks at m/e 196/197 and (b) central cleavage
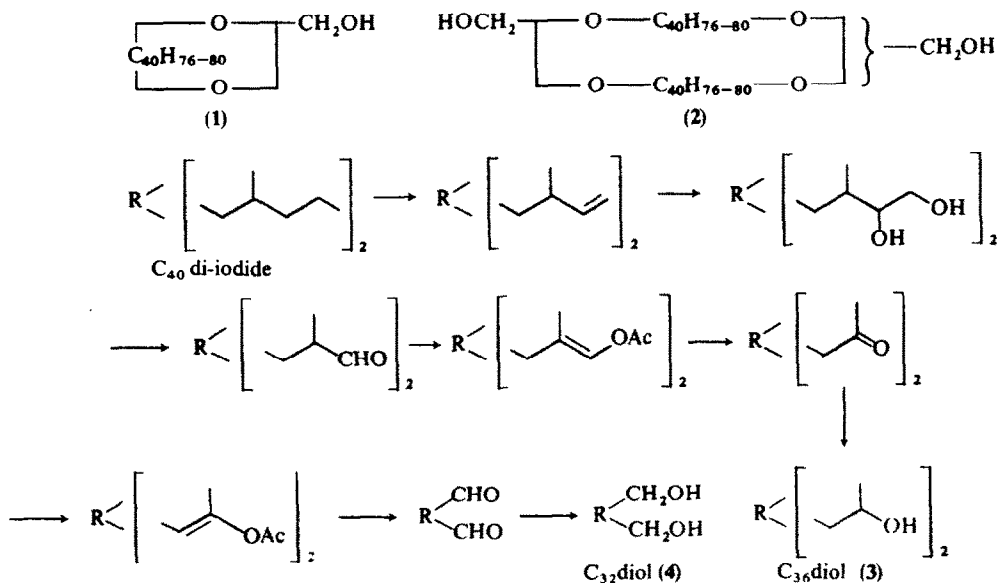
Scheme 1.
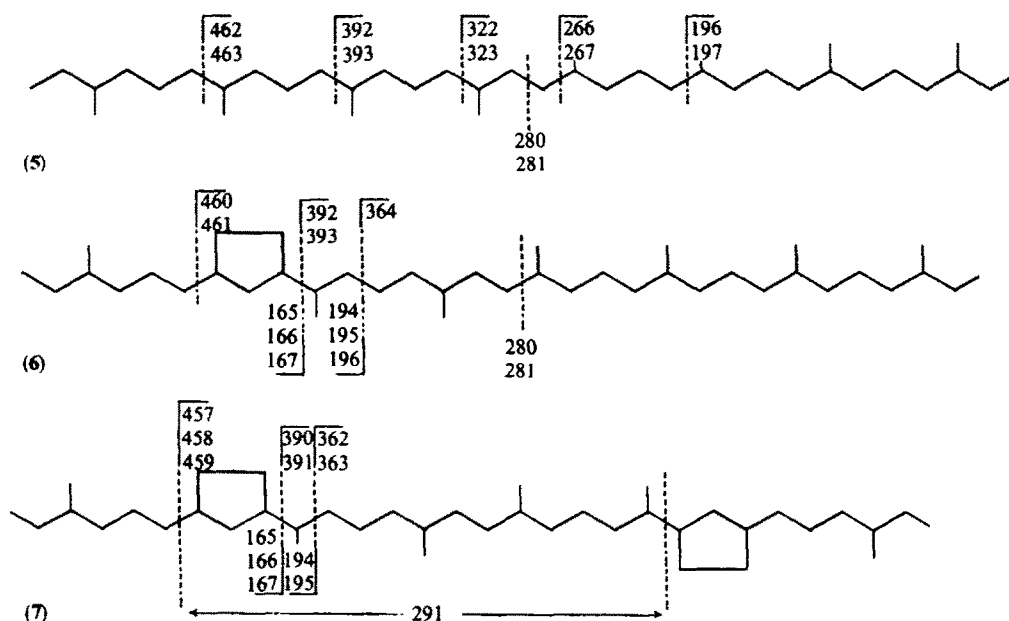
Scheme 2.

(again with or without H transfer), β to two $>$CHMe groups, to fragments at m/e 280,281. The two types of fragmentation are independent since in chemical ionization spectra (kindly provided by C. J. W. Brooks) only the molecular ion and the base peak from series a appear. All the fragmentation peaks were shifted by 1 mass unit in the spectra of the $^2H_2$-hydrocarbon. With the NMR evidence that the corresponding functionalized derivatives [e.g. the diol $C_{40}H_{80}(OH)_2$] contain 8 Me groups (see below), and the biogenetic evidence for an isoprenoid skeleton [5,7], the data require structure (5) for the hydrocarbon $C_{40}H_{82}$, as in Scheme 2. In particular they provide strong

evidence for the central head-to-head linkage in the ω,ω'-biphytanyl skeleton.

The MS of the monocyclic hydrocarbon $C_{40}H_{80}$ (m/e 560 and m/e 562 in the $^2H_2$-material) shows significant differences. While the molecular ion and the first major fragments at m/e 460,461 have one formal unsaturation the next fragments, at m/e 392 and 393, do not. Cleavage at the same bond also generates a group of strong peaks at m/e 165–167 (base peak, 166). There are corresponding cleavages apparently at an adjacent bond, differing by 28 mass units; the 'central' cleavage gives only fragments of m/e 280 and 281, presumably because the other moiety, for which peaks at m/e 278

and 279 would be expected, generates instead the smaller fragments at m/e ca 166 and ca 194. As before all the fragmentation peaks were shifted by 1 mass unit in the spectrum of $^2H_2$-material. Since the NMR evidence (below) shows that in the corresponding monocyclic diol there are just 7 CHMe groups we may conclude that the eighth Me group of the parent skeleton is used in forming the ring; this leads to structure (6) and the rationalization of the MS data shown in Scheme 2.

Finally for the bicyclic hydrocarbon $C_{40}H_{78}$ the molecular ion m/e 558 (560 in the $^2H_2$-material) and the fragments at m/e 457–459, 390–391, and 362–363 (all shifted by 1 mass unit in the spectra of the $^2H_2$-material) show a further deficit of 2 mass units when compared with the data for (6), the important counterparts at m/e 165–166 and 194–195 appearing as before. The next major fragment in the spectrum has m/e 291 corresponding to a C_{21} fragment with two formal unsaturations. This peak is *not* shifted in the spectrum of the $^2H_2$-hydrocarbon; it is therefore formed by cleavage at each end of the molecule and moreover it retains only one of the two rings originally present. Again, the NMR evidence (below) shows that in the corresponding diol etc. there are only six methyl groups, all secondary, which in the europium-expanded spectrum give three separated doublets, implying that they are symmetrically disposed in the molecule. This leads to structure (7) for the bicyclic hydrocarbon and the rationalization shown in Scheme 2 for its MS fragmentation.

NMR data: C_{40} diols

High-resolution NMR measurements (300 MHz; $CDCl_3$) of a tetraether sample in which the bicyclic C_{40} component predominated clearly indicated that in this component there are six $>$CHMe groups in the molecule. However, in order to obtain definitive data it was clearly essential to investigate the separate C_{40} components; preparative GLC of the C_{40} di-iodides was impossible because of their thermal instability but the di-acetates were successfully separated by this technique, and converted into the separate C_{40} diols: $C_{40}H_{80}(OH)_2$ (acyclic), $C_{40}H_{78}(OH)_2$ (monocyclic), and $C_{40}H_{76}(OH)_2$ (bicyclic). The relevant data obtained by NMR measurements using the europium shift reagent are summarized in Table 1.

With additions of up to 1:1 mole ratio of $Eu(fod)_3$ the signals for protons up to the fourth carbon from

Table 1. Values of δ (from Me_4Si) for $CDCl_3$ and for 1:1 mole ratio Eu (fod-d_9)$_3$ in $CDCl_3$ for the C_{4m} diols (9), (10), and (11)

Assignment* (protons on C no.)	δ (CDCl³)	δ (Eu shifted)	Present in (integral)
1,1'	3.67	8.87	9,10,11 (4H)
2,2'	1.30	4.70, 4.30†	9,10,11 (4H)
3,3'	1.55	3.65	9,10,11 (2H)
4,4'	1.28	2.58. 2.18†	9,10,11 (4H)
17.17'	0.90	2.15	9,10,11 (6H)
18.18'	0.85	0.97	9 (6H), 10 (3H)

*Numbering as in formulae (9–11); assignments supported by integration, multiplicity, and decoupling.
†Methylene protons non-identical with respect to adjacent chiral $>$CHMe.

the alcohol oxygen were fully resolved, and showed that in all three C_{40} diols there are two identical end-groups —$CH_2CHMeCH_2CH_2OH$. It is convenient to use the convention accepted for polyprenols and designate these C_5 units as $\alpha\alpha'$, and similarly, inwards from the end-groups the remainder as $\beta\beta'$, $\gamma\gamma'$, and $\delta\delta'$. The spectra obtained for the acyclic diol also showed a resolved signal corresponding to two further CHMe groups (in the $\beta\beta'$ units) at 0.85 shifting to 0.97 at 1:1 mole ratio; the same signal, but of half the intensity (i.e. for only one CHMe) appeared in spectra of the monocyclic diol but was completely absent in the spectra obtained with the bicyclic diol. Thus the cyclizations involve the methyl carbons of the β units; the β and β' units are identical in the acyclic diol, different in the monocyclic, and again identical in the bicyclic diol. Similarly in the NMR spectra of the C_{40} diacetates measured in C_6D_6 the acyclic diacetate shows three separate doublets, at δ 0.83 (6H), 0.92 (6H) and 0.95 (12H) assigned to the $\alpha\alpha'$, $\beta\beta'$ and ($\gamma\gamma'$ + $\delta\delta'$) units respectively; the bicyclic diacetate shows δ 0.83 (6H), no signal at 0.92, and a pair of doublets at 0.94 and 0.96—i.e. the CHMe of the β and β' units are missing and the $\gamma\gamma'$ and $\delta\delta'$ units are now somewhat differentiated.

NMR data: C_{36} and C_{32} bicyclic diols

Further data were obtained from NMR shift studies on the C_{36} and C_{32} bicyclic diols obtained from the C_{40} bicyclic diacetate by the route shown in Scheme 1 (see Experimental), which incidentally provides direct chemical evidence for the identical α and α' units. The NMR results are summarized in Tables 2 and 3.

Table 2. Values of δ (from Me_4Si) for $CDCl_3$ and for 1:1 mole ratio Eu(fod-d_9)$_3$ in $CDCl_3$ for the bicyclic C_{36} diol (3)

Assignment* (protons on C no.)	δ (CDCl³)	δ (Eu shifted)	Integral
3,3'	3.00	14.00	2H
17,17'	1.16	5.96	6H
4,4'	1.40	7.40, 6.20†	4H
5,5'	1.28	5.18	4H
6,6'	1.28	4.03	4H
7,7'	1.70	3.20	2H

*Numbering as in formulae 9–11; assignments supported by integration, multiplicity, and decoupling.
†Methylene protons non-identical with respect to adjacent chiral $>$CHMe

Table 3. Values of δ (from Me_4Si) for $CDCl_3$ and for 1:1 mole ratio Eu(fod-d_9)$_3$ in $CDCl_3$ for the bicyclic C_{32} diol (4)

| Assignment* (protons on C no.)	δ (CDCl³)	δ (Eu shifted)	Integral
4,4'	3.63	9.7	4H
5,5'	1.38	5.2	4H
6,6'	1.28	4.1	4H
7,7'	1.70	3.3	2H
19,19'	0.85	0.95	6H
20,20'	0.85	0.85	6H

*Numbering as in formulae 9–11; assignments supported by integration, multiplicity, and decoupling.

In the case of the C_{36} diol the original spectrum shows resolved signals for the residues of the $\alpha\alpha'$ units, —CHOHMe, at δ 1.16 (C$\underline{H}_3$) and 3.00 (C$\underline{H}$); with up to 1:1 molar ratio of Eu(fod)$_3$ these signals were progressively shifted to 5.96 (C$\underline{H}_3$) and 14.00 (C$\underline{H}$) and simultaneously the spectra showed resolved signals for one pair of C$\underline{H}$ and three further pairs of C$\underline{H}_2$ groups, which are assigned to the ring structure. With *ca* 3:1 of Eu(fod)$_3$: diol the remaining four Me groups could be distinguished as doublets at 1.02 and 0.85, assigned to CHMe of the $\gamma\gamma'$ and $\delta\delta'$ units. Similar results were obtained with the bicyclic C_{32} diol; in this case the $\gamma\gamma'$ and $\delta\delta'$ CHMe signals were clearly separated with a 1:1 molar ratio of Eu(fod)$_3$. Taken together the NMR data indicate part-structure (8) for the C_{40} bicyclic diol, and with the MS data for the C_{40} hydrocarbons (5)–(7) this leads to structures (9) (10) and (11) for the acyclic, monocyclic, and bicyclic diols.

W, 80–100 mesh operating at 310° with N$_2$ carrier at 180 ml min^{-1}. Column chromatography was on Merck Si gel, and TLC on Merck F254 Si gel plates. IR spectra were measured in liquid films (Perkin-Elmer 257), optical rotations in CHCl$_3$ (Perkin-Elmer 115). Petrol is bp 40–70°.

Tetraether and diacetate. The mixed tetraethers, obtained as described [4], were separated by TLC in CHCl$_3$–MeOH (9:1) into the less polar diglycerol tetraethers, R_f 0.95, and the more polar glycerol nonitol tetraethers, R_f 0.35. The latter had MW 1490 (C$_{92}$H$_{168-176}$O$_{12}$ requires 1464–1472). The diglycerol tetraethers had MW 1290(C$_{86}$H$_{164-172}$O$_6$ requires 1292–1300); acetylation (Ac$_2$O, 5 ml; C$_5$H$_5$N, 0.5 ml; 3 hr reflux) gave the mixed diglycerol tetraether diacetate, R_f 0.8 in petrol-Et$_2$O (3:1); [α]$_D$ + 6.6° (*c*, 2.0); MW 1380 (C$_{90}$H$_{168-176}$O$_8$ requires 1376–1384); v_m 1740, 1240 cm^{-1}, δ 4.14 (4H, *m*, CH$_2$OAc), 3.5 (14H, *m*, C$\underline{H}_2$-O and C$\underline{H}$-O), 2.05 (6H, *s*, C$\underline{H}_3$-CO).

Cleavage of the tetraethers. The diglycerol tetraether mixture (100 mg) was treated with BCl$_3$ (5 ml in 5 ml CHCl$_3$, 18°, 12 hr); the reaction mixture was evapd under N$_2$ and chromatographed; CHCl$_3$ eluted the C$_{40}$ dichlorides (93 mg) already described [4].

(8)

(9)

(10)

(11)
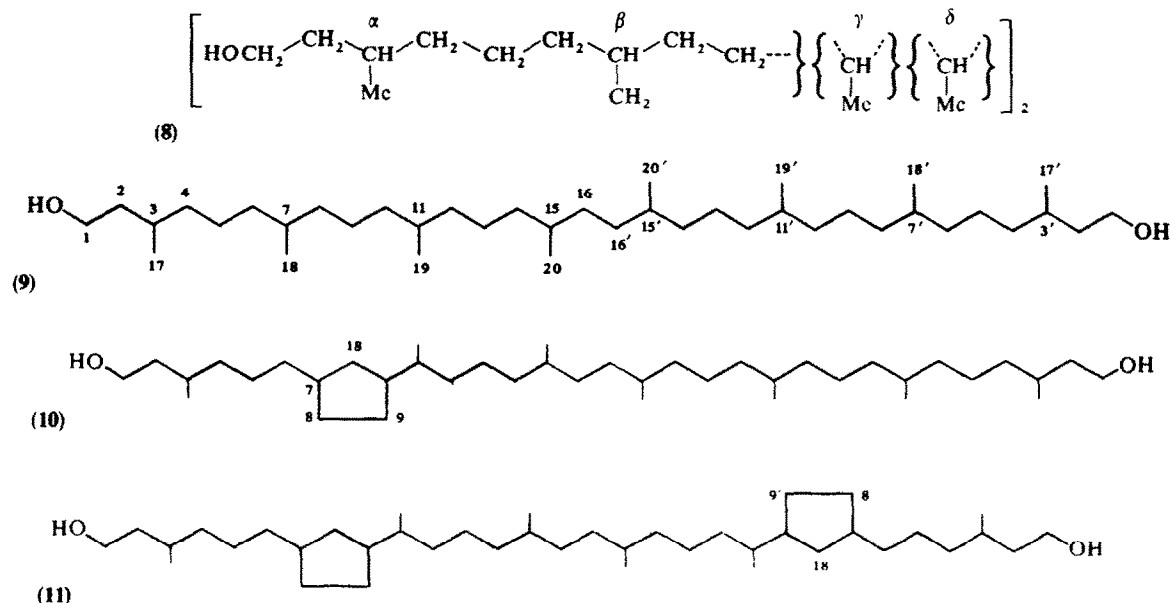
EXPERIMENTAL

General. Isolation and culture methods for the MT strains of Caldariella were as described [2], using the MT-3 strain grown at 75°. The isolation and physical data for the mixed tetraethers (previously called diethers) and the derived C$_{40}$ di-iodides have also been reported in full [4].

Instrumental. Some ^{1}H NMR (300 MHz) spectra were run at Manchester on the Varian SC-300; most were carried out at Arco Felice using the Varian XL-100-15; solns were normally in CDCl$_3$ with tetramethylsilane. MS were determined with the A.E.I. MS 30 at 70 eV. For GC-MS work the instrument was connected to a glass column, 1.5 m × 3 mm, packed with 1% OV-1 on GasChrom (100–120 mesh), temp. programmed at 6° min^{-1} over 220–310°, carrier gas He at 60 ml min^{-1}. For chemical ionization MS (Dr. C. J. W. Brooks, Glasgow) a Du Pont 21-490F instrument fitted with a dual electron impact/chemical ionization source was used; both types of spectrum could be recorded within 1 min of each other by admitting the reagent gas (isobutane). Optimal source temp. was 180°. GLC analyses of the C$_{40}$ hydrocarbons have already been described [4]. for preparative GLC. of the C$_{40}$ diacetates we used a Carlo Erba Fractovap GV equipped with a steel column 2m × 10mm. packed with 10% SE-30 on Chromosorb

and CHCl$_3$–MeOH (7:3) eluted glycerol (13 mg) (mol. ratio C$_{40}$ dichloride : glycerol 1.04).

C$_{40}$ hydrocarbons. Cleavage of the diglycerol tetraethers with HI followed by LiAlH$_4$ reduction gave the mixed C$_{40}$ hydrocarbons as previously described [4]. On GC-MS this product gave three peaks: C$_{40}$H$_{82}$, *m/e* 562(8%, M$^+$), 463(12), 462(11), 393(13), 392(11), 323(18), 322(19), 281(30), 280(16), 267(33), 266(32), 197(100); C$_{40}$H$_{80}$, *m/e* 560(14%, M$^+$), 461(3), 460(4), 393(11), 392(13), 364(7), 281(12), 280(15), 197(19), 196(27), 195(100), 194(94), 166(100), 165(75); C$_{40}$H$_{78}$, *m/e* 558(5%, M$^+$), 449(3) 458(2), 457(3), 391(6), 390(7), 363(2), 362(4), 291(10), 195(75), 194(71), 166(71) 165(100). In chemical ionization MS the major ions from the mixture were *m/e* 562(12%, M$^+$), 561(19), 560(19, M$^+$), 559(36), 558(31, M$^+$), 557(55), 555(12), and 195(100). For the di-deuterated hydrocarbons, see Results.

C$_{40}$ diols. The mixed di-iodides (2 g) were treated with AgOAc (4 g) (in AcOH, 300 ml; 24 hr reflux). Water was added and the mixture extracted with Et$_2$O; the Et$_2$O layer was washed successively with H$_2$O, aq. NaHCO$_3$, aq. Na$_2$S$_2$O$_3$, and H$_2$O, dried and evapd. The residue was chromatographed in petrol-Et$_2$O, and with 20% Et$_2$O the C$_{40}$ diacetates (1.4 g) were eluted. The mixed diacetates were separated by prep GLC

(recovery *ca* 60%) to give the acyclic (24 mg), monocyclic (112 mg), and bicyclic (630 mg) C_{40} diacetates (ν_m 1745, 1235, 1030 cm^{-1} for all three). Acyclic C_{40} diacetate [for numbering, cf. (9)] : $[\alpha]_D^{20}$ 0.00° (*c*, 1.0); *m/e* 678 (M$^+$, < 1%), 616 (M$^+$-AcOH, 2), 558(M$^+$-2AcOH, 2), 459 (M$^+$-2AcOH-C$_7$H$_{15}$, 2), 389(15), 319(12), 279(25), 195(100); δ(C$_6$D$_6$) 3.97 [4H, *t*(*J* = 7Hz), CH$_2$-CH$_2$-OAc], 1.92(6H. *s*, CH$_3$-CO), 1.55(*ca* 8H, *b*, CHMe), δ 0.95 [12H, *d*(*J* = 5.5 Hz), CH-CH$_3$ (19,19', 20,20')], 0.92 [6H, *d*(*J* = 5.5 Hz), CH-CH$_3$(18,18')], 0.83 [6H, *d*(*J* = 5.5 Hz), CH-CH$_3$ (17,17')]. Monocyclic C_{40} diacetate [for numbering, cf. (10)] : $[\alpha]_D^{20}$ − 13.3° (*c*, 1.0); *m/e* 676 (M$^+$, 2%), 616 (M$^+$-2AcOH, 2), 556 (M$^+$-2AcOH, 2), 517 (M$^+$-AcOH-C$_7$H$_{15}$, 2), 457 (M$^+$-2AcOH-C$_7$H$_{15}$, 2), 389(6), 223(35), 165(100); δ(C$_6$D$_6$) 3·97 [4H, *t*(*J* = 7 Hz), CH$_2$-CH$_2$-OAc], 1.92(6H, *s*, CH$_3$-CO), 1.80(*ca* 2H, *b*, ring CH), 1.55(*ca* 7 H, *b*, CHMe), 0.95[12 H, *d*(*J* = ·5.5 Hz), CH-CH$_3$ (19,19' 20,20')], 0.92 [3H, *d*(*J* = 5.5 Hz), CH-CH$_3$ (18')], 0.83 [6H, *d*(*J* = 5.5 Hz), CH-CH$_3$ (17,17')]. Bicyclic C_{40} diacetate [for numbering, cf. (11)] : $[\alpha]_D^{20}$ + 5.2°(*c*, 2.0); *m/e* 674 (M$^+$, 2%), 614 (M$^+$-AcOH, 2), 554 (M$^+$-2AcOH, 2), 515 (M$^+$-AcOH-C$_7$H$_{15}$, 2), 455 (M$^+$-2AcOH-C$_7$H$_{15}$, 2), 291(5), 223(40), 165(100); δ (C$_6$D$_6$) 3.97 [4 H, *t*(*J* = 7 Hz), CH$_2$-CH$_2$-OAc], 1.92 (6 H, *s*, Me-CO), 1.80 (*ca* 4H, *b*, ring CH), 1.55(*ca* 6H, *b*, CHMe), 0.96 [6H, *d*(*J* = 5.5 Hz), CH-CH$_3$(20,20')], 0.94 [6H, *d*(*J* = 5.5 Hz CH-CH$_3$(19,19')], 0.83 [6H, *d*(*J* = 5.5 Hz), CH-CH$_3$(17,17')]. The separate diacetates were saponified (10% aq. KOH, 6 hr reflux); the hydrolyzate was diluted with H$_2$O and extracted several times with Et$_2$O to afford, separately, the C_{40} diols (9), (10), and (11); for ^{1}H NMR and Eu shift data cf. Table 1.

Bicyclic C_{36} diol (cf. Scheme 1). The bicyclic C_{40} diacetate (0.60 g) was treated with HI (50 ml of 57% HI; 24 hr reflux); the product taken up in petrol, was washed successively with H$_2$O, satd aq. K$_2$CO$_3$, aq. Na$_2$S$_2$O$_3$, H$_2$O, and H$_2$O–MeOH (1:9) and finally chromatographed in *n*-hexane. The bicyclic C_{40} di-iodide thus prepared (0.61 g) was treated with M KOBut in dry ButOH (8.5 ml) at 85°; after 12 hr the reaction mixture was diluted with aq. MeOH and extracted with petrol; prep TLC afforded the bicyclic C_{40} diene $C_{40}H_{74}$ (0.25 g), R_f 0.8 (in Et$_2$O–petrol (1:9) on AgNO$_3$-treated Si gel), R_t 20.3 min (on 1% OV-1 at 235°); $[\alpha]_D^{20}$-4.4° (*c*, 3.0); *m/e* 554 (M$^+$, 9%), 483(2), 470(3), 455(3), 329(3), 291(3), 205(7), 192(18), 191(18), 165(100); ν_m 1654, 1000 and 915 (—CH═CH$_2$) cm^{-1}; δ(CCl$_4$) 5.66–5.5 (2H, seven lines, CHMe—CH═CH$_2$), 4.94–4.84 (4H, 15 lines, CH═CH$_2$), 1.73(7–8H, *m*), 1.26(b), 0.97[6H, *d*(*J* = 7.5 Hz), MeCH—CH═CH$_2$], 0.85 [15 H, two *d*(*J* = 5.5, 6 Hz), CH—CH$_3$]. The diene (0.25 g) was stirred with OsO$_4$ (0.24 g in 10 ml dry C$_5$H$_5$N) at 18° for 5hr, then aq. NaHSO$_3$ (3 g in 20 ml) and C$_5$H$_5$N (3 ml) were added and stirred for 30 min. Extraction with CH$_2$Cl$_2$ afforded the bicyclic C_{40} tetrol (0.32 g), ν_m 3350 and 1050 cm^{-1} δ 3.5 (complex, CHOH and CH$_2$OH), which without purification was treated with Pb(OAc)$_4$ (0.43 g) in AcOH (8 ml) and MeOH 940 ml) at 18° for 4 hr. Normal work-up gave the C_{38} dialdehyde (0.32 g), ν_m 1710 cm^{-1}, (CCl$_4$) 9.48 *d*(*J* = 1.5 Hz) CH-CHO. The dialdehyde was refluxed for 3 hr in isopropenyl acetate (8 ml) containing conc H$_2$SO$_4$ (1 drop); the excess isopropenyl acetate was evapd under red. press. H$_2$O was added, and the whole extracted with Et$_2$O. The Et$_2$O extract, washed with H$_2$O, aq. Na$_2$CO$_3$,

and H$_2$O, dried and evapd, was chromatographed in petrol with up to 10% Et$_2$O, which eluted the C_{38} di-(enol acetate) (0.13 mg), ν_m 1750 and 1220 cm^{-1}. This was ozonised in EtOAc at −20°; evaporation of the solvent gave the crude C_{36} diketone (0.09 g), ν_m 1715 cm^{-1}; δ 2.30 [4H, *t*(*J* = 7Hz); CH$_2$-CH$_2$-CO], 2.02(6H, *s*, CH$_3$-CO), 0.88 [14–15H, *d*(*J* = 5 Hz), CH-CH$_3$]. Part of this diketone (20 mg) with LiAlH$_4$ (in dry Et$_2$O, 12 hr), worked up in the normal way, and chromatographed in petrol–Et$_2$O (2:3) gave the bicyclic C_{36} diol (3) (16 mg). Part of this was used for ^{1}H NMR and Eu shift measurements (Table 2) and part was acetylated (Ac$_2$O–C$_5$H$_5$N) to give the bicyclic C_{36} diacetate: *m/e* 618 (M$^+$, 2%), 558 (M$^+$-AcOH, 2), 498 (M$^+$-2AcOH, 427(3), 137(100); δ (C$_6$D$_6$) 4.78 (2H, *m* CHOAc), 1.91 (6H, *s*, CH$_3$COO), 1.15 [6H, *d*(*J* = 6 Hz), CH$_3$-CH-O], 0.96 [6H, *d*(*J* = 5.5 Hz), CH-CH$_3$ (20,20')], 0.94 [6H, *d*(*J* = 5.5 Hz), CH-CH$_3$ (19,19')].

Bicyclic C_{32} diol (4). Using the methods described above, the remainder of the C_{36} diketone (70 mg) treated with isopropenyl acetate gave the C_{36} di-(enol acetate) (20 mg) ν_m 1255 and 1215 cm^{-1}, ozonolysis of which gave the C_{32} dialdehyde (14 mg), ν_m 1710 cm^{-1}, δ 9.45 [*t*(*J* = 6Hz), CH$_2$-CHO], reduced with LiAlH$_4$ to the bicyclic C_{32} diol (4) (6 mg). Part of this was used for ^{1}H NMR and Eu shift measurements (Table 3) and the remainder acetylated to give the C_{32} diacetate; *m/e* 562(M$^+$, 2%), 502 (M$^+$-AcOH, 2), 442(M$^+$-2AcOH, 2), 399(2),109(100); δ(C$_6$D$_6$) 3.97 [4H, *t*(*J* = 7 Hz), CH$_2$-CH$_2$-OAc], 1.92 (6H, *s* CH$_3$-CO), 0.96[6H, *d*(*J* = 5.5 Hz), CH-CH$_3$ (20,20')], 0.94[6H, *d*(*J* = 5.5 Hz), CH-CH$_3$ (19,19')].

REFERENCES

1. de Rosa, M., Gambacorta, A., Millonig, G. and Bu'Lock, J. D. (1974) *Experientia* **30**, 860.
2. de Rosa, M., Gambacorta, A. and Bu'Lock, J. D. (1975) *J. Gen. Microbiol.* **86**, 156.
3. Millonig, G., de Rosa, M., Gambacorta, A. and Bu'Lock, J. D. (1975) *J. Gen. Microbiol.* **86**, 165.
4. de Rosa, M., Gambacorta, A. and Bu'Lock, J. D. (1976) *Phytochemistry* **15**, 143.
5. de Rosa, M., Gambacorta, A., Minale, L. and Bu'Lock, J. D. (1974) *Chem. Commun.* 543.
6. Langworthy, T. A. and Mayberry, W. R. (1976) *Soc. Gen. Microbiol. Proc.* **3**, 165.
7. de Rosa, M., de Rosa, S. and Gambacorta, A. (1977)*Phytochemistry* **16**, 1909.
8. Darland, G., Brook, T. D., Samsonoff, W. and Conti, S. F. (1970) *Science* **170**, 1416.
9. Kates, M., Yengoyan, L. S. and Sastry, P. S. (1965) *Biochim. Biophys. Acta* **98**, 252.