Total Synthesis of Calditol: Structural Clarification of this Typical Component of Archaea Order *Sulfolobales***

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Abstract: The original structure of calditol—that is, an open-chain branched nonitol—has recently been questioned by various research groups and cyclopentane-based structures have been proposed. To unambiguously clear up this confusion, four isomeric cyclopentane candidates 26-29 have been synthesized. Of these, compound 27 was found to be fully identical to the natural product present in *Sulfolobus solfataricus* (A.T.C.C. 49155). The synthesis of 27 uses a samariumdiiodide-induced pinacolization reaction of the ketoaldehyde 15 as the critical step.

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total synthesis

Keywords: archaea • calditol • structure elucidation • sulfolobales

Introduction

In addition to prokaryotes and eukaryotes, archae have been introduced, on the basis of 16S ribosomal RNA sequence analyses,^[1] as the third group of living organisms. *Sulfolobus* is a genus of sulfur-oxidizing archae characterized by aerobic growth at high temperatures and low pH in the presence of elemental sulfur.^[2] Organisms of this genus are usually found in sulfur-containing habitats such as acidic hot springs and mud holes. The known species of Sulfolobus include Sulfolobus acidocaldarius, originally discovered in Yellowstone National Park (USA) and Sulfolobus solfataricus, originally discovered in Pisciarelli (Italy), and these have been widely used in research. Brock et al. suggested that Sulfolobus may be important geochemical agents in the production of sulfuric acid from elemental sulfur in high-temperature hydrothermal systems.^[2a] The membranes of extreme thermoacidophiles are based on two major types of complex macrocyclic tetraethers in which two polyols are linked together through two isoprenoid chains.^[3] In one type, the hydrophilic portions are two glycerol units in which the ether bonds are located at the sn-2 and sn-3 positions of glycerol.^[4] The diglycerol

tetraether structure **1** has been termed diglycerocaldarchaeol^[5] or GDGT (glycerodialkylglycerol tetraether).^[6]

The second type is based on glycerol on one hand and on a polyol on the other, for which the trivial name calditol has been coined.^[7] It may thus be termed calditoglycerocaldarch-



aeol according to Nishihara et al.,^[5] or glycerodialkyl calditol tetraether (GDCT). The history of this polyol is very interesting.

The early history of calditol—Langworthy and De Rosa: In 1974, Langworthy et al.^[8] studied the structure of various lipids extracted from *Sulfolobus acidocaldarius* (Strain 98-3), an archae originally isolated by Brock et al.^[2] from Locomotive Spring in Yellowstone National Park. One component, identified as a glycolipid B, after acidic methanolysis to remove the glucose unit gave a product which was identified as the polyol dialkyl glycerol triether **2** (Scheme 1). Although



Scheme 1. Isolation of a polyol dialkyl glycerol triether 2 by Langworthy.

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the structure of this new polyol was not determined at that time, they suggested that it might be cyclic, and demonstrated that it was connected through an ether linkage to glycerol.

At about the same time, the Italian group of De Rosa studied the structure of the second lipid type from *Caldariella acidophila* (Strain MT-4), isolated from an acid hot spring in



Agnano, Naples.^[9] This archaeon should now be renamed^[2c] Sulfolobus solfataricus (A.T.C.C. 49155). In complete contrast to Langworthy, they proposed that calditol was a unique acyclic branched-chain nonitol **3** containing no ether bond, and the structure of the corresponding calditol glycerol tetraether **4** was proposed by the Italian group. The stereochemistry of the calditol was not determined.



An improved purification of this so-called glycerol dialkyl nonitol tetraether (GDNT) has subsequently been reported.^[10]

Structural and chemical investigations-doubts about the correctness of the original structure of calditol: In 1990, Jeganathan and Vogel reported the first synthesis of two defined stereoisomers of De Rosa's nonitol.^[11] The ¹H NMR spectra of the corresponding peracetates were both different from the one reported by De Rosa et al. for the natural calditol peracetate. They concluded at this stage that the natural product was another stereoisomer. In 1995, we reported the chemical synthesis of another defined peracetylated isomer.[12] Comparison of the 13C NMR skeletal chemical shifts of this synthetic compound with those given by De Rosa for the nonacetate of calditol revealed odd discrepancies. From a critical analysis of De Rosa's structural work, we detected various significant anomalies, so that we seriously questioned the correctness of the original structure assigned to this natural product by the Italian group.^[12]

In addition to these synthetic efforts, two independent structural investigations of calditol also invalidated the original open-chain structure. Sugaï et al.,^[13] in accordance with Langworthy's prediction and on the basis of NMR analysis, proposed the new structure **5** for calditol, which could be isolated in pure form from type two lipids of *Sulfolobus acidocaldarius* (A.T.C.C. 33909). Arigoni's group independently reinvestigated the structure of the calditol

Abstract in French: La structure d'origine du calditol—à savoir un nonitol branché en chain droite—a été sérieusement remise en question récemment par plusieurs groupes de recherche, et des structures cyclopentaniques ont été proposées. Afin de totalement clarifier cette situation plutôt confuse, quatre candidats isomères structuraux cyclopentaniques 26–29 ont été synthétisés. Seul l'un d'eux, le composé 27, est totalement identique au produit naturel présent chez Sulfolobus solfataricus (A.T.C.C. 49155). La synthèse de 27 met en jeu dans l'étape critique une réaction de pinacolisation du cétoal-déhyde 15, initiée par le diiodure de samarium.



tetraether lipids from *Sulfolobus solfataricus* and also detected several significant differences in comparison with the spectral data reported by De Rosa et al.^[14] The interpretation of NMR data of their own lipid sample led to the proposed new structure **6** for calditol. In both structures **5** and **6**, a cyclic polyol is connected to *sn*-1 glycerol by an ether linkage. These structures only differ in the configuration at a single stereocenter.

Results and Discussion

To clarify this rather confusing situation, we launched a programme of chemical synthesis of various stereoisomers. A key derivative in this piece of work was the protected Dglucopyranose derived diol 14. In this compound, the hexitol derivative is connected through an ether linkage to the sn-1 position of di-O-benzylated glycerol. The strategy was then to convert the diol into a substituted cyclopentane ring by pinacolization. The synthesis of the key diol 14 is given in Scheme 2. To alkylate the sn-1 position of glycerol, we first transformed the sn-3-O-benzyl glycerol (7)^[15] into the cyclic sulfate 9, via the cyclic sulfite 8. Sharpless demonstrated that cyclic sulfates are easily opened by nucleophilic attack at the less hindered position.^[16] Indeed, phenyl 2,3,6-tri-O-benzyl-1thio- β -D-glucopyranose (10)^[17] was efficiently alkylated^[18] with cyclic sulfate 9 to give, after acidic hydrolytic removal of the transient sulfate, the secondary alcohol 11. Benzylation of this product gave compound 12, which was converted into the hemiacetal 13 using N-bromosuccinimide in moist acetone, and then into the target intermediate 14 (in 84% yield from 12) after reduction with lithium aluminum hydride. This synthetic pathway is of general interest for the efficient anchoring of a glyceryl appendage onto a secondary alcohol. As shown in Scheme 3, the glucitol derivative 14 was submitted to Swern oxidation^[19] to afford the unstable ketoaldehyde 15 which was not isolated. Immediate samarium-diiodide-promoted pinacolization gave an inseparable mixture of two *cis* diols, **16** and **17**, in a 3:1 ratio (78% yield). A similar pinacolization reaction has recently been reported.[20]

We then moved on to the problem of the inversion of configuration at C-1 of **16** and **17**. Our first approach consisted of the transformation of the *cis* diols into the corresponding separable cyclic sulfates and their subsequent opening with a nucleophile by an $S_N 2$ mechanism. Several nucleophiles such as lithium acetate, cesium acetate, ammonium benzoate, and sodium azide were tried in various solvents (DMF, HMPT, acetonitrile), but only elimination products and/or starting material were detected and no opening of the sulfate was observed.



Scheme 2. Synthesis of glucitol derivative 14. i) SOCl₂, Et₃N, CH₂Cl₂; ii) NaIO₄, RuCl₃, 0°C, CH₂Cl₂; iii) a) nBuLi, THF, HMPA; b) 2M H₂SO₄; iv) BnBr, NaH, THF; v) NBS, dark, acetone/H₂O; vi) LiAlH₄, THF.



Scheme 3. Synthesis of cis diols 16 and 17 by pinacol coupling. i) oxalyl chloride, DMSO, Et₃N, CH₂Cl₂, -65°C; ii) SmI₂, tBuOH, THF, -40°C.

A strategy consisting of the oxidation of the alcohol at C-1, followed by the reduction of the resulting ketone to the inverted alcohol was then examined. For this purpose, the mixture of cis diols 16 and 17 were conveniently separated as their carbonates 18 and 19 (Scheme 4), which were then



Scheme 4. Protection of the cis diols 16 and 17 as carbonates 18 and 19 and acetonides 20 and 21. i) 1,1'-carbonyl diimidazole, CH2Cl2; ii) K2CO3, CH₃OH; iii) 2,2 dimethoxypropane, acetone, CSA.

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carbonate in 87-92% yield to afford the pure cis diols 16 and 17, respectively. The structures of the two cyclopentane rings were unambiguously established at this stage by recording NOESY spectra of the corresponding acetonides 20 and 21 (Figure 1, Table 1). The discriminating nuclear Overhauser effects observed were as follows: 1) irradiation of H-6 of acetonide 20 gave an NOE with H-1 and H-4 indicating that H-1 and H-4 are on the same (top) face of the cyclopentane ring, and 2) irradiation of one of the methyl groups of the acetonide

21 gave an NOE with H-4 and H-2 indicating that H-4, H-2 and the hydroxyl groups of the corresponding diol 17 are all on the same (top) face of the cyclopentane ring. Thus compound 16 has the diol moiety located under the cyclopentane ring, whereas compound 17 has the diol unit pointing above the cyclopentane ring.



Figure 1. Nuclear Overhauser effects observed for acetonides 20 and 21. Benzyl groups (R) are not shown for clarity.

Table 1. Nuclear Overhauser effects observed for acetonides 20 and 21.

Irradiated proton ^[a]	NOE observed	
	Acetonide 20	Acetonide 21
CH ₃ (isopropylidene)	H-1, H-6	H-4 (weak), H-6, benzyl
CH ₃ (isopropylidene)	H-3	H-2 (weak), H-4
H-1	H-2	H-4, H-6
H-2	H-1, benzyl	benzyl
H-3	H-2, H-4, CH ₃	H-4, H-6
H-4	H-1, H-3 (weak)	-
H-6, H-6'	H-1, H-4, benzyl	H-4, CH ₃

[a] Key NOEs are indicated in bold.

The oxidation of the cis diols 16 and 17 was problematic. Swern oxidation^[19] gave a mixture of elimination products, whereas the Dess-Martin periodinane^[21] cleaved the C-C bond formed during the pinacol coupling to afford the glucitol derivative 14 after sodium borohydride reduction. Bromine oxidation of the diol-derived stannylenes, followed by sodium borohydride reduction, also gave poor results. Finally, and as observed by Adinolfi et al. in a similar case,^[22] oxidation was achieved in a satisfying yield using sodium hypochlorite/ 2,2,6,6-tetramethylpiperidine-N-oxide (TEMPO) methodology.^[23] Subsequent sodium borohydride reduction of the resulting ketones 22 and 23, probably directed by the tertiary alcohol, selectively afforded the expected trans diols 24 (Scheme 5) and 25 (Scheme 6). Traces of cis diols were easily separated after conversion into their corresponding acetonides 20 and 21, respectively. Catalytic hydrogenolysis of the trans diols 24 and 25 afforded the corresponding heptitols, which were directly peracetylated to give compounds 26 and 27, respectively. The same sequence was applied to the cis diols 16 and 17 to afford the peracetylated compounds 28 and 29, respectively (Scheme 5 and 6). Comparison of the ¹H and 13 C NMR spectra of the peracetates 26–29 with an authentic sample of peracetylated calditol, kindly provided to us by A. Sugaï, showed that the peracetylated natural product and compound 27 were identical. Furthermore, the optical rotations of both synthetic 27 and natural calditol heptacetate were very close (natural sample: $[\alpha]_{\rm D} = -14.6$ (c = 1.56 in CHCl₃), synthetic compound 27: $[\alpha]_D = -14$ (c = 0.27 in $CHCl_3)).$

Conclusion

This synthetic work unambiguously demonstrates for the first time that the structure of calditol (including its absolute configuration) isolated from the archaeon Sulfolobus acid-



Scheme 5. Synthesis of peracetylated compounds 26 and 28. i) TEMPO, NaOCl, CH₂Cl₂, H₂O; ii) NaBH₄, CH₃OH; iii) H₂, 10% Pd/C, EtOAc, CH₃OH then Ac₂O, pyridine, DMAP.





De Rosa or Arigoni for direct comparison, we consider it highly probable that the calditol present in various archaea has the same structure. This work also firmly establishes that the cyclic part of calditol is connected through an ether linkage to the sn-1 carbon atom of glycerol. It is worth noting that kerufarrides^[24] and crasserides,^[25] which are natural products of marine origin, contain a cyclopentane-pentol moiety that is also connected through an ether linkage, probably to sn-1 position of glycerol.

same archaeon that had been extensively studied by De Rosa

et al. and initially named Caldariella acidophila. Although no

sample of peracetylated calditol has been available to us from

Experimental Section

Cyclic sulfite 8: Diol 7 (9.0 g, 49.4 mmol) and Et₃N (27.7 mL, 198 mmol) were dissolved in anhydrous CH2Cl2 (100 mL), and the solution was cooled to 0°C. A solution of freshly distilled SOCl₂ (5.4 mL, 74.1 mmol) in anhydrous CH2Cl2 (50 mL) was added dropwise under argon. The reaction mixture was then stirred at RT for 1 h, diluted with cold Et₂O (100 mL), and washed with cold water (2 × 100 mL) and brine (100 mL). The organic

phase was dried over MgSO4 and filtered, and the solvent was removed in vacuo. Purification by column chromatography (EtOAc/cyclohexane 1:6 then 1:4) afforded cyclic sulphite 8 (8.0 g, 78%). This compound was immediately used in the next step.

Cyclic sulfate 9: H₂O (120 mL), Ru- $Cl_3 \cdot 3H_2O$ (1 mg), and then $NaIO_4$ (15 g, 70 mmol) were added to a solution of 8 (8.0 g, 38.5 mmol) in CH₃CN (80 mL) and CH₂Cl₂ (80 mL) at 0°C. The reaction mixture was stirred at 0 °C for 45 min, diluted with diethyl ether (150 mL), and the aqueous phase was extracted with Et2O $(2 \times 100 \text{ mL})$. Organic extracts were combined, washed with 5% aq Na2- S_2O_3 solution (2 × 100 mL), and dried over MgSO₄. The solvent was removed under reduced pressure to afford 9 (8.42 g, 89%) as a brown oil. MS (CI, NH₃): m/z (%): 245 (15) $[M+H]^+$, 262 (100) $[M+NH_4]^+$; elemental analysis calcd (%) for C₁₀H₁₂O₅S (244.3): C 49.17, H: 4.95; found: C 49.54, H 4.82

Alcohol 11: Compound 10 (15 g, 27.7 mmol) and 2,2'-bisquinoline (20 mg) were dissolved in anhydrous THF (150 mL). The solution was cooled to -40° C, and *n*BuLi (17.4 mL, 10.9 mmol, 1.6 M in hexane) was added dropwise until the solution became orange. Hexamethyl phosphoramide (HMPA; 30 mL) was

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ocaldarius (A.T.C.C. 33909) by Sugaï et al.^[13] is that shown in Figure 2.

The depicted relative stereostructure is indeed that previously proposed by Gräther and Arigoni, working on a sample isolated from Sulfolobus solfataricus (A.T.C.C. 49155), the



Figure 2. Structure of calditol.

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added dropwise under stirring followed by a solution of cyclic sulfate 9 (6.7 g, 27.7 mmol) in anhydrous THF (10 mL). The reaction mixture was allowed to warm to RT and acidified to pH 1 with H₂SO₄ (2 M). The reaction mixture was then heated at reflux for 3 h, cooled to RT, neutralized with solid NaHCO₃, filtered, diluted with Et₂O (200 mL), and washed with water (200 mL). The aqueous layer was extracted with Et₂O (2×200 mL). Organic extracts were combined and dried over MgSO4, and the solvent removed under reduced pressure. Purification by column chromatography (EtOAc/cyclohexane 1:4 then 1:2) afforded alcohol 11 (17 g, 87%) as a white solid. M.p. $60-61^{\circ}C$; $[\alpha]_{D}^{22} = +3.8 \ (c = 0.51 \text{ in CHCl}_{3})$; ¹H NMR $(CDCl_3, 400 \text{ MHz}): \delta = 7.64 - 7.30 \text{ (m}, 25 \text{ H}; 5 \text{ Ph}), 4.97 \text{ (d}, J = 10.7 \text{ Hz}, 2 \text{ H};$ 2 CHPh), 4.91 (d, J=10.7 Hz, 1H; CHPh), 4.78 (d, J=10.2 Hz, 1H; CHPh), 4.72 (d, J = 9.7 Hz, 1 H; H-1), 4.69 (d, J = 12.0 Hz, 1 H; CHPh), 4.63 (d, J = 12.0 Hz, 1 H; CHPh), 4.57 (s, 2 H; CH₂Ph), 3.94 (m, 1 H; H-8), 3.90 -3.82 (m, 3H; H-6, H-6', H-7), 3.78 (dd, J = 3.8 Hz, 10.2 Hz, 1H; H-7'), 3.72 (appt, J = 8.9 Hz, 1H; H-3), 4.02 (appt, J = 9.3 Hz, 1H; H-4), 3.57 (appt, J=9.5 Hz, 1H; H-2), 3.51 (m, 1H; H-5), 3.49 (app d, J=5.3 Hz, 2H; H-9, H-9'), 3.05 (d, J = 4.3 Hz, 1H; OH); ¹³C NMR (CDCl₃, 100 MHz): $\delta =$ 137.99, 137.89, 137.84, 137.83, 133.58 (5 $C_{\rm ipso}$), 131.86 – 127.41 (5Ph), 87.32 (CH-1), 86.14 (CH-3), 80.69 (CH-2), 78.92 (CH-5), 78.37 (CH-4), 75.67 (CH₂Ph), 75.26 (CH₂Ph), 74.21 (CH₂Ph), 73.36 (CH₂Ph), 73.29 (CH₂Ph), 70.92 (CH2-9), 69.93 (CH-8), 68.84 (CH2-6); MS (CI, NH3): m/z (%): 724 (100) $[M+NH_4]^+$; elemental analysis calcd (%) for $C_{43}H_{46}O_7S$ (706): C 73.06, H 6.56; found: C 73.05, H 6.65.

Thiophenyl derivative 12: A solution of alcohol 11 (9 g, 12.7 mmol) in anhydrous THF (80 mL) was added dropwise to a cooled suspension of BnBr (1.8 mL, 15.1 mmol) and NaH (624 mg, 15.6 mmol, 60 % dispersion in mineral oil) in anhydrous THF (30 mL). The solution was stirred at RT for 3 h, cooled to 0°C, and quenched with CH₃OH (100 mL). The solvent was removed under reduced pressure, and the residue dissolved in Et2O (200 mL), and washed with water (3×200 mL). The organic layer was dried over MgSO₄, and the solvent was removed under reduced pressure. Purification by column chromatography (EtOAc/cyclohexane 1:8 then 1:7) afforded **12** (9.43 g, 93 %) as a yellow oil. $[\alpha]_{D}^{22} = -2.1$ (c = 0.93 in CHCl₃); ¹H NMR (C₆D₆, 400 MHz): $\delta = 7.82 - 7.30$ (m, 30 H; 6 Ph), 5.03 (d, J =10.7 Hz, 1H; CHPh), 4.99 (s, 2H; CH₂Ph), 4.81 (d, J = 10.7 Hz, 1H; CHPh), 4.73 (d, J=9.1 Hz, 1H; H-1), 4.63 (s, 2H; CH₂Ph), 4.56 (d, J= 12.0 Hz, 1 H; CHPh), 4.51 (d, J = 12.0 Hz, 1 H; CHPh), 4.43 (s, 2 H; CH₂Ph), 4.16 (dd, J=5.2, 9.8 Hz, 1H; H-7), 3.91 (dd, J=4.6, 9.8 Hz, 1H; H-7'), 3.82-3.76 (m, 3H; H-6, H-6', H-8), 3.70-3.60 (m, 5H; H-2, H-3, H-4, H-9, H-9'), 3.31 (m, 1H; H-5); ¹³C NMR (C_6D_6 , 100 MHz): $\delta = 139.95$, 139.93, 139.66, 139.55, 139.45, 135.49 (6C_{ipso}), 132.65, 129.61, 129.04-127.90 (6Ph), 88.32 (CH-1), 87.40 (CH-2), 81.69 (CH-4), 79.93 (CH-5), 79.08 (CH-3), 78.57 (CH-8), 75.93 (CH₂Ph), 75.90 (CH₂Ph), 73.93 (CH₂Ph), 73.91 (CH₂Ph), 73.37 (CH₂-7), 72.69 (CH₂Ph), 71.08 (CH₂-9), 69.74 (CH₂-6); MS (CI, NH₃): m/z (%): 724 (100) $[M+NH_4]^+$; elemental analysis calcd (%) for C₅₀H₅₂O₇S (797.03): C 75.34, H 6.57; found: C 75.31, H 6.60.

Alcohol 13: Compound 12 (5.9 g, 7.4 mmol) was dissolved in acetone/water (290 mL, 95% v/v). The reaction mixture was cooled to 0°C and NBS (6.6 g, 36.9 mmol, recrystallized from water) was added in one portion in the dark. After 3 min of vigorous stirring, sat. aq Na2CO3 solution (200 mL) was added, and a white precipitate formed. The acetone was evaporated, and the remaining aqueous white suspension was extracted with CH₂Cl₂/ cyclohexane (2×200 mL, 50 % v/v). Organic extracts were combined and dried over $MgSO_4$, and the solvent removed under reduced pressure. The resulting solid was recrystallized from cyclohexane to afford 13 (4.6 g, 89%) as a white solid. Data for the major α anomer: ¹H NMR (CDCl₃, 400 MHz): $\delta = 7.37 - 7.30$ (m, 25 H; 5 Ph), 5.23 (app t, J = 3.0 Hz, 1 H; H-1), 4.92 (d, J = 10.8 Hz, 1 H; CHPh), 4.84 (d, J = 10.8 Hz, 1 H; CHPh), 4.79 (d, 12.1 Hz, 1 H; CHPh), 4.59 (d, J = 12.1 Hz, 1 H; CHPh), 4.55 (d, J = 12.2 Hz, 1 H; CHPh), 4.51 (s, 2 H; CH₂Ph), 4.49 (d, J = 12.2 Hz, 1 H; CHPh), 4.01 (m, 2H; H-5, H-7), 3.90 (t, J = 9.2 Hz, 1H; H-3), 3.72-3.65 (m, 4H; H-6, H-6', H-7', H-8), 3.56 (dd, J=3.0 Hz, J=9.2 Hz, 1H; H-2), 3.54 (m, 2H; H-9, H-9'), 3.48 (t, J = 9.2 Hz, 1H; H-4), 2.87 (d, J = 2.2 Hz, 1H; OH); ¹³C NMR (CDCl₃, 100 MHz): $\delta = 138.63$, 138.60, 137.94, 137.90, 137.81 (5 C_{ipso}), 128.46-127.51 (5Ph), 91.26 (CH-1), 81.46 (CH-3), 79.77 (CH-2), 78.16 (CH-4), 77.45 (CH-8), 75.47 (CH₂Ph), 73.35 (CH₂Ph), 73.31 (CH₂Ph), 73.21 (CH₂Ph), 72.53 (CH₂-7), 72.04 (CH₂Ph), 70.26 (CH-5), 70.31 (CH₂-9), 68.64 (CH₂-6); MS (CI, NH₃): *m/z* (%): 722 (100) [*M*+NH₄]⁺; elemental analysis calcd (%) for C44H48O8 (704.86): C 74.97, H 6.86; found: C 74.89, H 6.80.

D-Glucitol derivative 14: Alcohol 13 (5.1 g, 7.3 mmol) was dissolved in anhydrous THF (80 mL) under argon, the solution was cooled to 0 °C, and LiAlH₄ (0.55 g, 14.6 mmol) was added portionwise. The reaction mixture was allowed to warm to RT over 2 h. Ethyl acetate (80 mL) was added, and the solution acidified to pH1 with 1M HCl. The aqueous phase was extracted with EtOAc $(3 \times 80 \text{ mL})$, the organic extracts were combined. dried over MgSO₄, and concentrated. Purification by column chromatography (EtOAc/cyclohexane 1:5 then 1:2) afforded diol 14 (4.9 g, 95 %) as a colorless oil. $[\alpha]_D^{22} = +12.5$ (c = 0.56 in CHCl₃); ¹H NMR (CDCl₃, 400 MHz): $\delta = 7.37 - 7.34$ (m, 25 H; 5 Ph), 4.64 - 4.53 (m, 10 H; 5 CH₂Ph), 4.02 (m, 1H; H-5), 3.96 (dd, *J* = 4.4, 5.9 Hz, 1H; H-3), 3.82 (m, 2H; H-1, H-2), 3.77 (m, 5H; H-1', H-4, H-7, H-7', H-8), 3.69 (dd, J = 4.0, 9.7 Hz, 1H; H-6), 3.62 (dd, J = 5.3, 9.7 Hz, 1H; H-6'), 3.61 (m, 2H; H-9, H-9'), 3.35 (d, J = 6.1 Hz, 1H; OH-5), 2.37 (appt, 1H; OH-1); ¹³C NMR (CDCl₃, 100 MHz): $\delta = 138.18$, 138.14, 138.02, 138.00, 137.91 (5 C_{inso}), 128.35 -127.55 (5Ph), 79.61 (CH-4), 79.38 (CH-2), 79.30 (CH-3), 77.27 (CH-8), 74.50 (CH₂Ph), 73.31 (CH₂Ph), 73.26 (CH₂Ph), 72.69 (CH₂Ph), 72.06 (CH₂Ph), 71.63 (CH₂-7), 71.04 (CH₂-6), 70.32 (CH-5), 69.61 (CH₂-9), 61.62 (CH₂-1); MS (CI, NH₃): *m/z* (%): 724 (100) [*M*+NH₄]⁺; elemental analysis calcd (%) for $C_{44}H_{50}O_8$ (706): C 74.87, H 6.99; found: C 74.77, H 7.05.

Cis diols 16 and 17: Oxalyl chloride (2.3 mL, 26.7 mmol) was dissolved in anhydrous THF (180 mL) under argon. The solution was cooled to -78 °C, anhydrous DMSO (2.3 mL, 32.0 mmol) was added dropwise, and the solution stirred for 10 min at -60 °C. A solution of compound 14 (3.8 g, 5.3 mmol) in anhydrous THF (60 mL) was then added slowly, and the resulting solution stirred for 10 min at -60 °C. The reaction mixture was allowed to warm to $-45\,^{\circ}\text{C}$ and stirred for 45 min. Anhydrous Et₃N (7.5 mL, 53.4 mmol) was then added, the solution was stirred for 10 min and allowed to warm up to RT for 1 h. The resulting white suspension was then transferred by cannula under argon to a 1_M solution of SmI₂ in THF (60 mL) containing degassed tert-butanol (1 mL) at -78 °C. The reaction mixture was stirred for 4 h at -40° C and then allowed to warm up to RT. A semi-saturated aq. NH₄Cl solution (80 mL) was added to the reaction mixture, and the aqueous phase was extracted with EtOAc (3×80 mL). Organic extracts were combined and dried over MgSO4, and the solvent removed under reduced pressure. Purification by column chromatography (EtOAc/cyclohexane 1:4 then 1:1) afforded an inseparable 3:1 mixture of the cis diols 16 and 17 (2.9 g, 78%) as a yellow oil.

Carbonates 18 and 19: Diols 16 and 17 (2.4 g, 3.4 mmol) and 1,1'-carbonyl diimidazole (2.2 g, 13.75 mmol) were dissolved in anhydrous CH2Cl2 (150 mL) under argon. The solution was heated at reflux for 16 h, cooled to RT, and washed with water (2×100 mL). The organic phase was dried over MgSO4, and the solvent removed under reduced pressure. Purification by column chromatography (EtOAc/cyclohexane 1:7 then 1:5) afforded carbonate **19** (490 mg, 19%) as a colourless oil. $[\alpha]_{D}^{22} = -10.0$ (c = 0.55 in CHCl₃); ¹H NMR (C₆D₆, 400 MHz): $\delta = 7.41 - 7.20$ (m, 25 H; 5 Ph), 4.82 (dd, J=1.8, 3.5 Hz, 1 H; H-1), 4.76 (d, J=12.0 Hz, 1 H; CHPh), 4.72 (d, J= 12.0 Hz, 1H; CHPh), 4.59 (m, 3H; 3CHPh), 4.41 (m, 3H; 3CHPh), 4.26 (app d, J = 12.1 Hz, 2H; CH₂Ph), 3.89 (m, 2H; H-2, H-3), 3.82 (dd, J = 4.4, 5.7 Hz, 1H; H-4), 3.75 (m, 2H; H-7, H-7'), 3.72 (d, J = 11.2 Hz, 1H; H-6), 3.71 (m, 1H; H-8), 3.58 (m, 2H; H-9, H-9'), 3.51 (d, *J* = 11.2 Hz, 1H; H-6'); ¹³C NMR (CDCl₃, 100 MHz): $\delta = 153.87$ (C=O), 138.25, 137.99, 137.67, 137.18, 136.82 (5 C_{ipso}), 128.65-127.51 (5 Ph), 87.57 (C-5), 86.82 (CH-4), 83.85 (CH-3), 82.56 (CH-2), 82.18 (CH-1), 76.82 (CH-8), 73.64, 73.41, 73.03, 72.26, 72.11 (5 CH₂Ph), 71.84 (CH₂-7), 69.37 (CH₂-9), 68.11 (CH₂-6); MS (CI, NH₃): m/z (%): 748 (100) [M+NH₄]⁺; elemental analysis calcd (%) for C45H46O9 (730.8): C 73.95, H 6.34; found: C 73.78, H 6.52.

Further elution afforded carbonate **18** (1.7 g, 67%) as a colourless oil. $[\alpha]_{22}^{22} = +30.1$ (c = 1.02 in CHCl₃); ¹H NMR (C₆D₆, 400 MHz): $\delta = 7.44 - 7.19$ (m, 25 H; 5Ph), 4.76 (d, J = 11.7 Hz, 1H; CHPh), 4.68 (d, J = 12.0 Hz, 1H; CHPh), 4.64 (d, J = 12.0 Hz, 1H; CHPh), 4.63 (d, J = 11.7 Hz, 1H; CHPh), 4.59 (d, J = 11.7 Hz, 1H; CHPh), 4.51 (d, J = 11.7 Hz, 1H; CHPh), 4.59 (d, J = 11.7 Hz, 1H; CHPh), 4.51 (d, J = 5.4 Hz, 1H; H-1), 4.45 (s, 2 H; CH₂Ph), 4.35 (appt, J = 8.7 Hz, 1H; H-3), 4.30 (d, J = 11.7 Hz, 1H; CHPh), 4.26 (d, J = 12.0 Hz, 1H; CHPh), 4.15 (d, J = 12.0 Hz, 1H; CHPh), 3.94 (m, 2 H; H-7, H-7'), 3.83 (m, 1H; H-8), 3.73 (d, J = 8.6 Hz, 1H; H-4), 3.67 (app d, J = 4.6 Hz, 2H; H-9, H-9'), 3.61 (dd, J = 5.4, 8.7 Hz, 1H; H-2), 3.36 (d, J = 10.1 Hz, 1H; H-6), 3.38 (d, J = 10.1 Hz, 1H; H-6'); ¹³C NMR (C₆D₆, 100 MHz): $\delta = 154.52$ (C=O), 139.88, 139.51, 139.43, 138.41, 138.07 (5 C_{lpso}), 129.27 - 128.12 (5 Ph), 85.96 (C-5), 85.39 (C-3), 81.55 (CH-4), 79.91 (CH-2), 78.36 (CH-8), 77.52 (CH-1), 74.12 (CH₂Ph), 74.01 (CH₂Ph), 73.93 (CH₂Ph), 72.66 (CH₂Ph), 72.64 (CH₂-7), 72.59 (CH₂Ph), 70.58 (CH₂-9),

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70.34 (CH₂-6); MS (CI, NH₃): m/z (%): 748 (100) [M+NH₄]⁺; elemental analysis calcd (%) for C₄₅H₄₆O₉ (730.8): C 73.95, H 6.34; found: C 73.83, H 6.50.

Acetonide 20: Camphorsulphonic acid (3 mg) and 2,2-dimethoxypropane (2 mL, 16 mmol) were added to a solution of diol 16 (207 mg, 0.29 mmol) in acetone (18 mL) under argon. The solution was stirred at RT for 17 h, and the reaction mixture then neutralized with Et₃N. The solvent was removed in vacuo and purification by column chromatography (EtOAc/cyclohexane 1:5) afforded acetonide 20 (175 mg, 80%) as a colorless oil. $[\alpha]_{D}^{20} = -10.8$ $(c = 1.0 \text{ in CHCl}_3)$; ¹H NMR $(C_6D_6, 400 \text{ MHz})$: $\delta = 7.50 - 7.19 \text{ (m, 25H;}$ 5Ph), 5.03 (d, J=12.0 Hz, 2H; CH₂Ph), 4.81 (m, 3H; 3CHPh), 4.71 (dd, J = 7.5, 8.9 Hz, 1 H; H-3), 4.63 (d, J = 4.1 Hz, 1 H; H-1), 4.62 (d, J = 11.9 Hz, 1H; CHPh), 4.48 (s, 2H; CH₂Ph), 4.33 (app q, 2H; CH₂Ph), 4.15 (dd, J = 4.8, 9.7 Hz, 1H; H-7), 4.04 (app q, *J* = 4.8 Hz, 1H; H-8), 3.97 (dd, *J* = 4.1, 8.9 Hz, 1H; H-2), 3.94 (d, J = 7.3 Hz, 1H; H-4), 3.84 (dd, J = 4.8, 9.7 Hz, 1H; H-7'), 3.81 (dd, J=4.5, 10.1 Hz, 1H; H-9), 3.73 (dd, J=4.5, 10.1 Hz, 1 H; H-9'), 3.53 (d, J = 9.5 Hz, 1 H; H-6), 3.49 (d, J = 9.5 Hz, 1 H; H-6'), 1.69, 1.44 (2s, 6H; 2CH₃); ¹³C NMR (C₆D₆, 100 MHz): δ = 138.80, 139.59, 139.67, 140.21, 140.22, (5 C_{ipso}), 129.17-127.90 (5 Ph), 112.74 (C(CH₃)₂), 87.20 (CH-3), 85.24 (C-5), 83.39 (CH-4), 80.76 (CH-1), 80.21 (CH-2), 78.65 (CH-8), 74.22 (CH₂-6), 74.10 (CH₂Ph), 73.88 (CH₂Ph), 73.59 (CH₂Ph), 72.79 (CH₂Ph), 72.32 (CH₂Ph), 71.79 (CH₂-7), 71.67 (CH₂-9), 27.55, 27.59 (2 CH_3) ; MS (CI, NH₃): m/z (%): 762 (100) $[M+NH_4]^+$; elemental analysis calcd (%) for C47H52O8 (744.9): C 75.78, H 7.03; found: C 75.71, H 7.16.

Acetonide 21: This compound was synthesized from 17 by the procedure described for **20** and was obtained as a colorless oil. $[\alpha]_{D}^{22} = +6.2$ (c = 0.7 in CHCl₃); ¹H NMR (C₆D₆, 400 MHz): $\delta = 7.45 - 7.19$ (m, 25 H; 5 Ph), 4.96 (d, J = 12.0 Hz, 1 H; CHPh), 4.94 (d, J = 2.5 Hz, 1 H; H-1), 4.92 (d, J = 12.0 Hz, 1H; CHPh), 4.83 (d, J = 12.0 Hz, 1H; CHPh), 4.73 (d, J = 12.0 Hz, 1H; CHPh), 4.68 (d, J = 12.0 Hz, 1 H; CHPh), 4.62 (d, J = 12.0 Hz, 1 H; CHPh), 4.49 (dd, *J* = 7.1, 9.5 Hz, 1 H; H-3), 4.50–4.41 (m, 4 H; 2 CH₂Ph), 4.35 (dd, J = 2.5, 7.1 Hz, 1 H; H-2), 4.32 (d, J = 9.5 Hz, 1 H; H-4), 4.19 (dd, J = 4.3, 10.3 Hz, 1 H; H-7), 4.00 (dd, J = 5.1, 10.3 Hz, 1 H; H-7'), 3.97 (d, J = 10.3 Hz, 1 H; H-6), 3.91 (m, 1 H; H-8), 3.90 (d, J = 10.3 Hz, 1 H; H-6'), 3.71 (m, J = 5.3 Hz, 2H; H-9, H-9'), 1.57, 1.52 (2s, 6H; 2CH₃); ¹³C NMR (C₆D₆, 100 MHz): $\delta = 140.01, 139.84, 139.55, 139.35, 139.11$ (5 C_{ipso}), 129.02 – 127.84 (5Ph), 113.68 (C(CH₃)₂), 90.87 (CH-4), 88.39 (C-5), 87.46 (CH-4), 86.15 (CH-2), 84.57 (CH-1), 78.46 (CH-8), 74.15 (CH₂Ph), 73.90 (CH₂Ph), 73.37 (CH₂Ph), 72.71 (CH₂Ph), 72.26 (CH₂Ph), 72.00 (CH₂-7), 71.03 (CH₂-9), 70.72 (CH₂-6), 29.16, 27.43 (2CH₃); MS (CI, NH₃): m/z (%): 762 (100) [M+NH₄]⁺; elemental analysis calcd (%) for C₄₇H₅₂O₈ (744.9): C 75.78, H 7.03; found: C 75.67, H 7.15.

Cis diol 16: Carbonate 18 (545 mg, 0.75 mmol) was dissolved in CH₃OH (50 mL). Potassium carbonate (309 mg, 2.24 mmol) was added under argon, and the reaction mixture stirred at RT. After 20 h, the reaction mixture was stirred with ion exchange resin IR-120 (2 g) for 30 min and then filtered. The solvent was removed under reduced pressure, and the residue was purified by column chromatography (EtOAc/cyclohexane 1:2) to afford the *cis* diol **16** (505 mg, 98% yield) as a colorless oil. $[\alpha]_{D}^{22} = +7.4$ $(c = 0.51 \text{ in CHCl}_3)$; ¹H NMR $(C_6D_6, 400 \text{ MHz})$: $\delta = 7.43 - 7.19 \text{ (m, 25H;}$ 5Ph), 4.69 (d, J=11.8 Hz, 1H; CHPh), 4.67 (s, 2H; CH₂Ph), 4.66 (d, J= 11.0 Hz, 1 H; CHPh), 4.62 (d, J = 11.0 Hz, 1 H; CHPh), 4.52 (d, J = 11.8 Hz, 1H; CHPh), 4.44 (s, 2H; CH₂Ph), 4.43 (m, 1H; H-3), 4.42 (d, J = 12.0 Hz, 1 H; CHPh), 4.35 (d, J = 12.0 Hz, 1 H; CHPh), 4.32 (dd, J = 6.1, 8.1 Hz, 1 H; H-1), 4.11 (d, J=6.0 Hz, 1 H; H-4), 4.02 (dd, J=4.5, 10.2 Hz, 1 H; H-7), 3.94 (m, 2H; H-2, H-7'), 3.81 (m, 1H; H-8), 3.66 (app d, J = 5.2 Hz, 2H; H-9, H-9'), 3.58 (d, J = 9.4 Hz, 1 H; H-6), 3.53 (d, J = 9.4 Hz, 1 H; H-6'), 3.52 (s, 1 H; OH-5), 3.19 (d, J = 8.1 Hz, 1 H; OH-1); ¹³C NMR (C₆D₆, 100 MHz): $\delta \,{=}\, 139.80, \, 139.54, \, 139.41, \, 139.24, \, 139.13, \, (5\,C_{ipso}), \, 129.08 \,{-}\, 128.14 \, \, (5\,Ph),$ 88.47 (CH-3), 83.76 (CH-4), 82.22 (CH-2), 78.12 (C-5), 78.11 (CH-8), 74.01 (CH₂Ph), 73.93 (CH₂Ph), 73.02 (CH₂Ph), 72.81 (CH₂Ph), 72.66 (CH₂Ph), 72.60 (CH2-6), 71.76 (CH2-7), 71.53 (CH-1), 70.88 (CH2-9); MS (CI, NH3): m/z (%): 722 (100) $[M+NH_4]^+$; elemental analysis calcd (%) for $C_{44}H_{48}O_8$ (704.8): C 74.97, H 6.86; found: C 74.85, H 7.00.

Cis diol 17: This compound was synthesized from carbonate 19 by the procedure described for diol 16 and was obtained as a colorless oil. $[a]_{D}^{22} = +9.0 \ (c = 0.1 \ \text{in CHCl}_3); {}^{1}\text{H} \text{ NMR } (C_6D_6, 400 \text{ MHz}): \delta = 7.48 - 7.17 \ \text{(m, 25 H; 5 Ph)}, 4.91 \ \text{(d, } J = 11.9 \text{ Hz}, 1 \text{ H}; \text{ CHPh}), 4.76 \ \text{(s, 2 H; CH}_2\text{Ph}), 4.69 \ \text{(d, } J = 11.9 \text{ Hz}, 1 \text{ H}; \text{ CHPh}), 4.64 \ \text{(d, } J = 2.1 \text{ Hz}, 2 \text{ H}; \text{ CH}_2\text{Ph}), 4.44 \ \text{(s, 2 H}; \text{CH}_2\text{Ph}), 4.30 \ \text{(d, } J = 11.9 \text{ Hz}, 2 \text{ H}; \text{CH}_2\text{Ph}), 4.26 \ \text{(app t, } J = 4.5 \text{ Hz}, 1 \text{ H}; \text{H-1}), 4.15 \ \text{(d, } J = 5.4 \text{ Hz}, 1 \text{ H}; \text{H-4}), 4.11 \ \text{(app t, } J = 5.4 \text{ Hz}, 1 \text{ H}; \text{H-4})$

1 H; H-3), 3.93 (dd, J = 4.4, 10.4 Hz, 1 H; H-7), 3.89 (dd, J = 5.0, 10.4 Hz, 1 H; H-7'), 3.88 (d, J = 9.5 Hz, 1 H; H-6), 3.80 (m, 1 H; H-8), 3.71 (d, J = 9.5 Hz, 1 H; H-6'), 3.65 (app d, J = 5.4 Hz, 2 H; H-9, H-9'), 3.40 (s, 1 H; OH-5), 2.90 (d, J = 4.4 Hz, 1 H; OH-1); ¹³C NMR (C₆D₆, 100 MHz): $\delta = 139.87$, 139.61, 139.58, 139.47, 138.84, (5 C_{ipso}), 129.15 – 128.10, (5 Ph), 88.28 (CH-4), 87.94 (CH-2), 84.40 (CH-3), 79.11 (C-5), 78.36 (CH-8), 77.58 (CH-1), 74.18 (CH₂Ph), 73.92 (CH₂Ph), 72.70 (CH₂-6, CH₂Ph), 72.60 (CH₂Ph), 72.53 (CH₂Ph), 71.56 (CH₂-7), 70.97 (CH₂-9); MS (CI, NH₃): m/z (%): 722 (100) [M+NH₄]⁺, 290 (15); elemental analysis calcd (%) for C₄₄H₄₈O₈ (704.8): C 74.97, H 6.86; found: C 74.91, H 6.98.

Trans diol 24: Diol **16** (36 mg, 0.05 mmol) was dissolved in CH_2Cl_2 (1 mL) and potassium bromide (4 mg, 0.034 mmol), TEMPO (40 mg, 0.25 mmol), and water (50 µL) were added. The solution was cooled to 0°C and a large excess of NaOCI (0.2 mL, technical solution diluted with water and adjusted to pH 9 just before use) was added dropwise. Some more NaOCI and TEMPO were added to complete the reaction. After 50 min, the reaction mixture was diluted with CH_2Cl_2 (5 mL) and washed with saturated aq. Na₂S₂O₃ solution (5 mL) and water (5 mL). The organic phase was dried over MgSO₄, and the solvent removed under reduced pressure to afford the crude ketone **22** as an orange oil. The ketone **22** was then dissolved in CH_3OH (5 mL), the solution was cooled to 0°C and NaBH₄ (6 mg, 0.15 mmol) was added. After 1 h, the solvent was removed under reduced pressure. Purification by column chromatography (EtOAc/cyclohexane 1:2) afforded the *trans* diol **24** (15 mg, 40%) as a colorless oil.

Data for cyclopentanone **22**: ¹H NMR (C₆D₆, 400 MHz): δ = 7.43 – 7.17 (m, 25 H; 5Ph), 5.18 (d, *J* = 11.8 Hz, 1 H; CHPh), 4.83 (d, *J* = 11.8 Hz, 1 H; CHPh), 4.75 (app d, *J* = 11.9 Hz, 2H; CH₂Ph), 4.51 (app d, *J* = 3.8 Hz, 2 H; CH₂Ph), 4.42 (m, 1H; H-4), 4.41 (m, 3H; 3 CHPh), 4.38 (d, *J* = 6.5 Hz, 1 H; H-2), 4.36 (d, *J* = 11.9 Hz, 1H; CHPh), 4.32 (dd, *J* = 0.8, 6.5 Hz, 1 H; H-3), 3.88 (dd, *J* = 4.5, 10.0 Hz, 1 H; H-7), 3.83 (d, *J* = 8.9 Hz, 1 H; H-6), 3.82 (dd, *J* = 4.5, 5.6 Hz, 1 H; H-7), 3.72 (s, 1 H; OH), 3.65 (m, 1 H; H-8), 3.61 – 3.55 (m, 3H; H-9, H-9', H-6'); ¹³C NMR (C₆D₆, 100 MHz): δ = 209.77 (C=O), 139.42, 139.31, 139.16, 138.74, 138.50, (5 C_{1ps0}), 129.13 – 128.25 (5 Ph), 85.06, 83.82, 80.78, 77.76 (CH-2, CH-3, CH-4, CH-8), 77.64 (C-5), 74.10, 73.98, 73.44, 73.40, 72.65, 72.42, 71.98, 70.25 (5 CH₂Ph, CH₂-6, CH₂-7, CH₂-9); MS (CI, NH₃): *m/z* (%): calcd for C₄₄H₄₉O₈ [*M*+NH₄]⁺: 720.3536; found: 720.3542.

Data for trans diol **24**: $[a]_D^{20} = +5.2$ (c = 0.3 in CHCl₃); ¹H NMR (C₆D₆, 400 MHz): $\delta = 7.49 - 7.18$ (m, 25H; 5Ph), 4.96 (d, J = 12.0 Hz, 1H; CHPh), 4.87 (d, J = 11.9 Hz, 1H; CHPh), 4.79 (d, J = 11.9 Hz, 1H; CHPh), 4.88 (d, J = 12.0 Hz, 1H; CHPh), 4.57 (s, 2H; CH₂Ph), 4.46 (app t, J = 5.8 Hz, 1H; H-1), 4.43 (s, 2H; CH₂Ph), 4.39 (d, J = 12.0 Hz, 1H; CHPh), 4.32 (app t, J = 7.0 Hz, 1H; H-3), 4.29 (d, J = 12.0 Hz, 1H; CHPh), 4.07 (m, 2H; H-2, H-4), 3.93 (dd, J = 4.5, 10.2 Hz, 1H; H-7), 3.85 (dd, J = 4.3, 10.2 Hz, 1H; H-7), 3.72 (s, 2H; H-6, H-6'), 3.70 (m, 1H; H-8), 3.63 (m, 2H; H-9, H-9'), 3.57 (s, 1H; OH-5), 2.98 (d, J = 5.6 Hz, 1H; OH-1); ¹³C NMR (C₆D₆, 100 MHz): $\delta = 139.74$, 139.72, 139.61, 139.32, 138.73 (5C_{ipso}), 129.15 – 128.07 (5Ph), 87.18 (CH-2), 86.01 (CH-3), 83.43 (CH-4), 83.09 (CH-1), 78.10 (C-5), 77.94 (CH-8), 74.25 (CH₂Ph), 73.96 (CH₂Ph), 73.67 (CH₂-6), 73.07 (CH₂Ph), 72.62 (CH₂Ph), 72.60 (CH₂Ph), 71.57 (CH₂-7), 70.57 (CH₂-9); MS (CI, NH₃): m/z (%): 722 (100) [M+NH₄]⁺, 290 (80); HRMS (CI +, NH₃): m/z calcd for C₄₄H₄₉O₈ [M+H]⁺: 705.3427; found: 705.3434.

Trans diol 25: This compound was obtained from diol 14 by the procedure described for diol 24 and was obtained as a colorless oil. $[\alpha]_{D}^{22} = +10.2$ (*c* = 0.56 in CHCl₃); ¹H NMR (CDCl₃, 400 MHz): $\delta = 7.40 - 7.30$ (m, 25 H; 5Ph), 4.72 - 4.53 (m, 10 H; 5CH₂Ph), 4.17 (t, *J* = 5.1 Hz, 1H; H-2), 4.04 (m, 2H; H-1, H-3), 3.93 (d, *J* = 9.7 Hz, 1H; H-6), 3.78 - 3.71 (m, 5H; H-4, H-6', H-7, H-7', H-8), 3.59 (m, 2H; H-9, H-9'), 3.03 (s, 1H; OH-5), 2.87 (d, *J* = 5.5 Hz, 1H; OH-1); ¹³C NMR (CDCl₃, 100 MHz): $\delta = 138.50$, 138.17, 138.03, 137.75, 137.73 (5 C_{ipso}), 128.45 - 127.49 (5Ph), 88.80 (C-4), 87.74 (C-3), 80.21 (C-5), 76.97 (C-8), 75.73 (CH-1), 73.67 (CH₂Ph), 73.32 (CH₂Ph), 72.23 (CH₂Ph), 72.09 (CH₂Ph), 71.97 (CH₂Ph), 70.83 (CH₂-7), 70.03 (CL₂-6), 70.05 (CH₂-9); MS (CI, NH₃): *m/z* (%): 722 (100) [*M*+NH₄]⁺; HRMS (CI +, NH₃): *m/z* calcd for C₄₄H₅₂NO₈ [*M*+NH₄]⁺: 722.3693; found: 722.3681.

Calditol isomer heptacetate 26: The diol **24** (7 mg, 0.01 mmol) was dissolved in CH₃OH (5 mL). Hydrogenolysis with 10% Pd/C was performed for 17 h by which time TLC (EtOAc/CH₃OH/H₂O 3:3:1) showed one non-UV active spot. The solution was filtered through Celite

and eluted with CH₃OH, and the solvent removed under reduced pressure. The residue was then dissolved in anhydrous pyridine (1 mL) under argon, and the solution cooled to 0°C. Ac₂O (0.5 mL) and DMAP (1 mg) were added. The reaction mixture was allowed to warm to RT and was stirred for 4 h. The solvent was then removed under reduced pressure and coevaporated with toluene $(3 \times 5 \text{ mL})$. Purification by flash chromatography (EtOAc/cyclohexane 1:1 then EtOAc) afforded the heptacetate 26 (4 mg, 73 %) as a colorless oil. $[\alpha]_{D}^{20} = +15.2 (c = 0.3 \text{ in CHCl}_{3}); {}^{1}\text{H NMR (CDCl}_{3},$ 400 MHz): $\delta = 5.80$ (m, 1H; H-1), 5.26 (m, 2H; H-2, H-3), 5.17 (m, 1H; H-8), 4.60 (d, J=12.0 Hz, 1 H; H-6), 4.49 (d, J=12.0 Hz, 1 H; H-6'), 4.33 (dd, J = 4.1, 11.9 Hz, 1H; H-7), 4.15 (dd, J = 6.4, 11.9 Hz, 1H; H-7'), 4.09(dd, J = 1.7, 2.7 Hz, 1 H; H-4) 3.90 (dd, J = 5.0, 10.3 Hz, 1 H; H-9), 3.64 (dd, J = 4.7, 10.7 Hz, 1H; H-9'), 2.15 (s, 3H; OAc), 2.14 (s, 3H; OAc), 2.13 (s, 3H; OAc), 2.12 (s, 3H; OAc), 2.11 (2s, 6H; 2OAc), 2.10 (s, 3H; OAc); ¹³C NMR (CDCl₃, 100 MHz): $\delta = 170.61$, 170.42, 170.19, 169.96, 169.74, 169.59, 169.47 (7 C=O), 83.02 (C-5), 81.72 (CH-4), 77.79 (CH-3), 76.54 (CH-1), 69.91 (CH-2), 69.05 (CH2-9), 62.37 (CH2-7), 61.81 (CH2-6), 21.24, 20.95, 20.84, 20.83, 20.77, 20.73, 20.71 (7 CH₃); MS (CI, NH₃): m/z (%): 722 (100) $[M+NH_4]^+$; HRMS (CI+, CH₄): m/z calcd for $C_{23}H_{33}O_{15}$ $[M+H]^+$: 549.1819; found: 549.1815.

Calditol heptacetate 27: This compound was obtained from diol **17** by the procedure described for compound **26** and was obtained as a colorless oil. $[a]_{D}^{20} = -14.4$ (c = 0.27 in CHCl₃); ¹H NMR (CDCl₃, 400 MHz): $\delta = 5.48$ (dd, J = 1.8, 4.6 Hz, 1H; H-1), 5.45 (dd, J = 4.6, 8.4 Hz, 1H; H-3), 5.37 (dd, J = 4.6, 8.4 Hz, 1H; H-2), 5.16 (m, 1H; H-8), 4.93 (d, J = 12.8 Hz, 1H; H-6), 4.53 (d, J = 12.8 Hz, 1H; H-6'), 4.36 (dd, J = 4.0, 11.8 Hz, 1H; H-7), 4.22 – 4.18 (m, 2H; H-4, H-7'), 3.81 (m, 2H; H-9, H-9'), 2.17 (s, 3H; OAc), 2.14 (s, 3H; OAc), 2.13 (s, 3H; OAc), 2.12 (s, 3H; OAc), 2.11 (s, 3H; OAc), 2.06 (s, 3H; OAc), 2.05 (s, 3H; OAc), 169.88, 169.75, 169.13 (7 C=O), 86.98 (C-5), 85.42 (CH-4), 80.21 (CH-3), 73.03 (CH-1), 72.78 (CH-2), 69.96 (CH-8), 69.94 (CH₂-7), 62.43 (CH₂-9), 58.73 (CH₂-6), 21.89, 20.94, 20.87, 20.73, 20.63, 20.59, 20.55 (7 CH₃); HRMS (CI +, CH₄): calcd for C₂₃H₃₃O₁₅ [M+H]⁺: 549.1819; found: 549.1823.

Calditol isomer heptacetate 28: This compound was synthesized from diol 16 by the procedure described for compound 26 and was obtained as a colorless oil. $[\alpha]_{D}^{20} = +17.9 (c = 0.7 \text{ in CHCl}_{3}); {}^{1}\text{H NMR} (\text{CDCl}_{3}, 400 \text{ MHz}):$ $\delta = 5.66 \text{ (dd, } J = 0.8, 5.4 \text{ Hz}, 1 \text{ H}; \text{H-1}\text{)}, 5.37 \text{ (dd, } J = 2.7, 6.2 \text{ Hz}, 1 \text{ H}; \text{H-3}\text{)},$ 5.25 (dd, J = 5.4, 5.9 Hz, 1 H; H-2), 5.19 (m, 1 H; H-8), 4.92 (d, J = 12.8 Hz, 1H; H-6), 4.33 (dd, J = 3.9, 11.8 Hz, 1H; H-7), 4.20 (d, J = 12.8 Hz, 1H; H-6'), 4.17 (dd, *J* = 6.9, 11.8 Hz, 1 H; H-7'), 3.89 (dd, *J* = 4.7, 10.1 Hz, 1 H; H-9), 3.81 (dd, J=0.6, 2.4 Hz, 1H; H-4), 3.56 (dd, J=4.3, 10.1 Hz, 1H; H-9'), 2.17 (s, 3H; OAc), 2.15 (s, 3H; OAc), 2.14 (s, 3H; OAc), 2.12 (s, 3H; OAc), 2.11 (s, 3H; OAc), 2.09 (s, 3H; OAc), 2.08 (s, 3H; OAc); ¹³C NMR $(CDCl_3, 100 \text{ MHz}): \delta = 170.66, 170.22, 170.17, 169.74, 169.69, 169.61,$ 169.57 (7 C=O), 83.09 (CH-4), 82.55 (C-5), 82.32 (CH-3), 73.18 (CH-2), 71.59 (CH-1), 69.98 (CH-8), 69.08 (CH2-9), 62.51 (CH2-7), 61.77 (CH2-6), 21.22, 20.95, 20.87, 20.71, 20.68, 20.49, 20.47 (7 CH₃C=O); elemental analysis calcd (%) for $C_{23}H_{32}O_{15}$ (548.5): C 50.36, H 5.88; found: C 50.26, H 6.02

Calditol isomer heptacetate 29: This compound was synthesized from diol **17** by the procedure described for compound **26** and was obtained as a colorless oil. $[\alpha]_{D}^{20} = -6.3$ (c = 0.35 in CHCl₃); ¹H NMR (CDCl₃, 400 MHz): $\delta = 5.59$ (d, J = 7.7 Hz, 1 H; H-1), 5.48 (ddd, J = 0.9, 4.5, 7.7 Hz, 1 H; H-2), 5.17 (m, 1 H; H-8), 5.12 (dd, J = 2.9, 4.5 Hz, 1 H; H-3), 4.66 (d, J = 12.1 Hz, 1 H; H-6), 4.62 (d, J = 12.1 Hz, 2 H; H-6, H-6'), 4.33 (m, 2 H; H-4, H-7), 4.17 (dd, J = 5.9, 11.9 Hz, 1 H; H-7'), 3.90 (dd, J = 5.0, 10.7 Hz, 1 H; H-9), 3.78 (dd, J = 4.5, 10.7 Hz, 1 H; H-9'), 2.14 (s, 3 H; OAc), 2.13 (s, 3 H; OAc), 2.11 (4s, 12 H; 4 OAc), 2.07 (s, 3 H; OAc); ¹³C NMR (CDCl₃, 100 MHz): $\delta = 170.55$, 170.28, 170.26, 169.94, 169.85, 169.69, 169.67 (7 C=O), 84.85 (C-5), 82.81 (CH-4), 78.25 (CH-2), 77.81 (CH-3), 75.52 (CH-1), 69.72 (CH-8), 69.21 (CH₂-9), 62.19 (CH₂-7), 61.85 (CH₂-6), 21.53, 20.86, 20.78, 20.72, 20.70, 20.66, 20.64 (7 CH₃C=O); elemental analysis calcd (%) for C₂₃H₃₂O₁₅ (548.5): C 50.36, H 5.88; found: C 50.20, H 5.96.

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